This article was downloaded by:

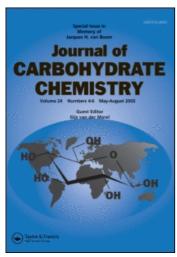
On: 23 January 2011

Access details: Access Details: Free Access

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

Chemical Synthesis of (4,6-Pyr)-Gal β1→4GlcNAcβ1→3Fucβ1→OMe: A Pyruvated Trisaccharide Related to the Cell Aggregation of the Sponge *Microciona Prolifera*

Shaojiang Deng^a; Biao Yu^a; Zhongwu Guo^a; Yongzheng Hui^a

^a State Key Laboratory of Bioorganic and Natural Products Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai, China

To cite this Article Deng, Shaojiang , Yu, Biao , Guo, Zhongwu and Hui, Yongzheng(1998) 'Chemical Synthesis of (4,6-Pyr)-Gal $\beta1$ —4GlcNAc $\beta1$ —3Fuc $\beta1$ —OMe: A Pyruvated Trisaccharide Related to the Cell Aggregation of the Sponge *Microciona Prolifera*', Journal of Carbohydrate Chemistry, 17: 3, 439 — 452

To link to this Article: DOI: 10.1080/07328309808002904 URL: http://dx.doi.org/10.1080/07328309808002904

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

CHEMICAL SYNTHESIS OF (4,6-Pyr)-Gal $\beta1\rightarrow 4$ GlcNAc $\beta1\rightarrow 3$ Fuc $\beta1\rightarrow 0$ Me: A PYRUVATED TRISACCHARIDE RELATED TO THE CELL AGGREGATION OF THE SPONGE MICROCIONA PROLIFERA

Shaojiang Deng, Biao Yu,* Zhongwu Guo, and Yongzheng Hui*

State Key Laboratory of Bioorganic and Natural Products Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, Shanghai 200032, China

Received July 20, 1997 - Final Form December 31, 1997

ABSTRACT

4,6-O-[(R)-1-carboxylethylidene]Gal $\beta1\rightarrow 4$ GlcNAc $\beta1\rightarrow 3$ Fuc $\beta1\rightarrow 0$ Me, a pyruvated trisaccharide unit involved in the aggregation factor of the marine sponge *Microciona prolifera*, was synthesized stereospecifically and unambiguously employing thioglycosides as glycosyl donors to construct glycosidic bonds.

INTRODUCTION

Species-specific reaggregation of dissociated cells from the marine sponge *Microciona prolifera* has been used for a long time as a simple model to study the molecular interactions involved in tissue development and organization of higher animals. Recent research has shown that the reaggregation of dissociated sponge cells mainly depends on a large extracellular adhesion proteoglycan, named marine cell aggregation factor (MAF). The multiple low affinity carbohydrate-carbohydrate interactions between the MAFs and the interactions between MAF and the proteoglycan on the outer integument of the cells form the basis for cell reaggregation in the sponge system. Two monoclonal antibodies, Block 1 and Block 2, raised against the native

Microciona aggregation factor can selectively inhibit the aggregation of the MAF.³⁻⁴ The corresponding carbohydrate epitopes of the MAF have been characterized to be a pyruvated trisaccharide $(4,6-Pyr)Gal\beta1\rightarrow 4GlcNAc\beta1\rightarrow 3Fuc$ for Block 1,³ and a sulfated disaccharide, $(3-SO_3Na)GlcNAc\beta1\rightarrow 3Fuc$, for Block 2.⁴ We have reported the chemical

synthesis of the sulfated disaccharide unit (2b).⁹ Recently, Ziegler reported a synthesis toward an aminopentyl trisaccharide of 1a.¹⁰ His synthesis employed lactose as a starting material to bypass the construction of the glycosidic bond between a pyruvated galactose and the 4-OH of a glucosamine unit. Herein, we wish to report a convergent synthesis of the methyl glycoside of this trisaccharide (1b) by using thioglycosides (8b, 14) as glycosyl donors to build the glycosidic bonds.

RESULTS AND DISCUSSION

The pyruvated phenyl thiogalactosides (8a and 8b) were prepared as outlined in Scheme 1. β-Phenyl thioglycoside 4,¹¹ readily prepared from β-D-peracetyl galactopyranose (3), was deacetylated followed by protection of the 4,6-OH with a benzylidene group to furnish the diol 5. Compound 5 was acetylated to give 6a, which was treated with 80% HOAc at 80 °C to give 7a with 4,6-OH free. Methyl pyruvate was then reacted with 7a to bridge the 4,6-OH groups according to Ziegler's method. However, treatment of the acetylated diol 7a with methyl pyruvate in the presence of BF₃OEt₂ (2 equiv) in CH₃CN resulted in very complex products and the expected 8a was isolated

Reagents and Conditions: (a) PhSH, BF₃-OEt₂, CH₂Cl₂, rt, 86%; (b) 1) MeONa, MeOH, rt; 2) PhCH(OMe)₂, DMF, TsOH, 50 °C, 85%; (c) Ac₂O, Py, rt, 92% for 6a; BzCl, Py, rt, 90% for 6b; (d) 80% HOAc, 80 °C, 95% for 7a; 80% for 7b; (e) CH₃COCOOMe, BF₃-OEt₂, MeCN, rt, 30% for 8a; 64% for 8b.

Scheme 1

only in a poor yield (30%). Therefore, benzoylated diol 7b was prepared from 5 and, as Ziegler reported, ¹³ converted into the desired 8b in good yield (64%).

The phenyl thioglycoside 14 was prepared as shown in Scheme 2. β-Phenyl thioglycoside 10, readily prepared from tetraacetate 9, was deacetylated and benzylidinated to give 11. Protection of the 3-OH of 11 with a benzyl group afforded 12, which was converted to 13 quantitatively by regioselective ring-opening of the benzylidene group by treatment with NaBH₃CN and HCl in ether. Compound 13 was then acetylated to give 14. The H NMR signal for 4-H of 14 was found to be shifted downfield to 5.12 ppm compared with that for 4-H of 13 (3.95 ppm).

Scheme 3 depicts construction of the target molecule (1b). 2,4-Di-O-benzyl- β -L-fucopyranoside (15), readily prepared according to our previous procedure,⁹ was efficiently glycosylated with phenyl thioglucoside 14 under promotion of NIS and AgOTf to generate the disaccharide 16 in 72% yield. The newly formed glycosidic bond was proved to be β -form from the coupling constant value ($J_{1',2'} = 8.2$ Hz). Originally, it was

Reagents and Conditions: (a) PhSH, BF₃-OEt₂, CH₂Cl₂, rt, 75%; (b) 1) MeONa, MeOH, rt; 2) PhCH(OMe)₂, DMF, TsOH, 50 °C, 80%; (c) BnBr, NaH, DMF, rt, 84%; (d) NaBH₃CN, THF, then HCl/Et₂O, rt, 100%; (e) Ac₂O, Py, rt, 100%.

Scheme 2

Reagents and Conditions: (a) NIS, AgOTf, CH_2Cl_2 , 4ÅMS, -20 °C to rt, 72%; (b) 1) NH₂NH₂, 95% EtOH, reflux; 2) Ac₂O, Py, rt; 3) NaOMe, MeOH, rt, 74%; (c) NaOMe, MeOH, rt, 100%; (d) 8b, NIS, AgOTf, CH_2Cl_2 , 4ÅMS, -20 °C to rt, 71%; (e) 1) NaBH₄, *i*-PrOH-H₂O, rt, then AcOH (pH 5), 80 °C; 2) Ac₂O, Py, rt; 3) NaOMe, MeOH, rt; 4) NaOH, H₂O, rt; 5) 10% Pd-C, H₂, 95% EtOH, 44%.

planned to convert the N-Phth to NHAc before introduction of the pyruvated galactose unit. Therefore, 4'-OH free disaccharide acceptor 17 was prepared by removal of the Phth group followed by N-acetylation and O-deacetylation. Unexpectedly, 17 was found to have very poor solubility in the usual solvents for glycosidation reactions, such as CH₂Cl₂, ClCH₂CH₂Cl, CH₃CN, THF, Et₂O, PhCH₃, etc.. Therefore, keeping the Phth protecting group intact, 16 was deacetylated to give 18. Compound 18, with 4'-OH free, was readily glycosylated with the pyruvated phenyl thiogalactoside 8b in the presence of NIS and AgOTf ^{15,16} to furnish the fully protected trisaccharide 19 in 71% yield. The final protecting group manipulation was successfully performed in a straightforward manner (6 steps, 44% yield). Compound 19 was treated with NaBH4 in i-PrOH and H2O followed by being heated in the acidified solution (acetic acid was added till pH to 5) at 80 °C to remove the Phth group. 17,18 The crude product was then N-acetylated, debenzoylated, saponified, and hydrogenolyzed sequentially to yield the target 1b. The structure of 1b was further ascertained from 1D ¹H NMR, DQFCOSY, and ESIMS spectral results. The three anomeric protons of 1b were determined to be at 4.86 ppm (H-1', $J_{1',2'}=8.5$ Hz), 4.53 ppm (H-1", $J_{1",2"}$ = 7.8 Hz), and 4.36 ppm (H-1, $J_{1,2}$ = 7.8 Hz).

In summary, the pyruvated trisaccharide, $4,6-O-[(R)-1-\text{carboxyethylidene}]Gal \beta 1 \rightarrow 4GlcNAc\beta 1 \rightarrow 3Fuc\beta 1 \rightarrow OMe (1b)$, which is related to the species-specific reaggregation of dissociated cells of *Microciona prolifera*, was unambiguously synthesized in a convergent and stereospecific manner.

EXPERIMENTAL

General methods. Melting points were uncorrected. Optical rotations were measured with a Perkin-Elmer Model 241 MC Polarimeter at 25 °C. TLC analyses were performed on precoated plates of Silica Gel HF₂₅₄ (0.5 mm, Qingdao, China) and detected by 10% sulfuric acid in MeOH. Flash column chromatography was performed on Silica Gel H (400 mesh). ¹H NMR spectra were recorded on a Bruker AM-300 or a Bruker AM-600 spectrometer with Me₄Si as an internal standard. IR spectra were recorded with a Shimadzu IR-440 spectrometer. Mass spectra were recorded on a VG QUATTRO or a HP5989A mass instrument.

Phenyl 4,6-*O*-Benzylidene-1-thio-β-D-galactopyranoside (5). To a solution of $4^{11.19}$ (4.49 g, 10.2 mmol) in MeOH (100 mL) and CH₂Cl₂ (100 mL) was added a catalytic amount of NaOMe. After being stirred at rt for 2 h, the mixture was neutralized with Dowex-50WX8 (H⁺ form), filtered, and concentrated *in vacuo*. The residue was dissolved in dry DMF (30 mL), then PhCH(OMe)₂ (3.1 mL) was added followed by camphorsulphonic acid (100 mg). After being stirred at 50 °C under diminished pressure for 2 h, the mixture was quenched with Et₃N (1mL), and extracted with EtOAc. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (CH₂Cl₂: MeOH 100:1 \rightarrow 50:1 \rightarrow 20:1) to afford 5^{20} (3.13 g, 85%): mp 119~120 °C; [α]_D -37.4° (*c* 1.2 CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.7-7.29 (m, 10H, 2C₆H₅), 5.52 (s, 1H, PhCH), 4.52 (d, 1H, H-1, J_{1,2} = 9.2 Hz), 4.40 (dd, 1H, H-6a, J_{6a,6b} = 12.5 Hz, J_{5,6a} = 1.1 Hz), 4.23 (m, 1H, H-4), 4.04 (dd, 1H, H-6b, J_{5,6b} = 1.5 Hz), 3.71 (m, 2H, H-2, H-3), 3.57 (m, 1H, H-5); IR (cm⁻¹): 3430 (OH), 1580, 1100-950; HREIMS: Cald for C₁₉H₂₀O₅S: 360.1032. Found: 360.1048.

Phenyl 2,3-Di-O-acetyl-4,6-O-benzylidene-1-thio-β-D-galactopyranoside (6a). A solution of 5 (137 mg, 0.38 mmol) in dry pyridine (2 mL) and Ac₂O (2 mL) was stirred overnight. The mixture was quenched with MeOH (1 mL), diluted with EtOAc. The organic layer was washed with 5% HCl solution, saturated NaHCO₃ solution, and brine respectively, and then dried over anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (petroleum ether:EtOAc 3:1) to afford $6a^{21}$ (147 mg, 92%) as a white amorphous solid: [α]_D +5.8° (c 0.33 CH₂Cl₂) [Lit²¹ +12° (c 1.0 CHCl₃)]; ¹H NMR (300 MHz, CDCl₃) δ 7.65-7.2 (m, 10H, 2C₆H₅), 5.47 (s, 1H, PhCH), 5.34 (t, 1H, H-2, J_{1,2} = J_{2,3} = 9.8 Hz), 5.00 (dd, 1H, H-3, J_{3,4} = 3.5 Hz), 4.71 (d, 1H, H-1), 4.37 (d, 1H, H-4), 4.37 (dd, 1H, H-6a, J_{6a,6b} = 12.5 Hz, J_{5,6a} = 1 Hz), 4.02 (dd, 1H, H-6b, J_{5,6b} = 1 Hz), 3.58 (m, 1H, H-5), 2.07, 2.02 (2s, 2×3H, 2CH₃CO); EIMS (*m/z*): 335 (M-PhS), 291, 275, 43 (base); IR (cm⁻¹): 1735, 1580, 1440, 1380, 1250, 1220, 1100, 1050.

Anal. Calcd for C₂₃H₂₄O₇S (444.50): C, 62.15; H, 5.44. Found: C, 62.11; H, 5.46.

Phenyl 2,3-Di-O-benzoyl-4,6-O-benzylidene-1-thio-β-D-galactopyranoside (6b). A procedure similar to that for the preparation of 6a was employed. Compound 5

(4.5 g) was treated with BzCl (6.3 mL) to afford **6b** (6.4 g, 90%): mp 169-170 °C (Lit²² 163 °C); $[\alpha]_D$ +64.2° (c 1.1 CHCl₃) [Lit²² +61.1° (c 1.2 CHCl₃)]; ¹H NMR (300 MHz, CDCl₃) δ 8.0-7.2 (m, 20H, 4C₆H₅), 5.81 (dd, 1H, H-2, J_{1,2} = 9.8 Hz, J_{2,3} = 9.9 Hz), 5.52 (s, 1H, PhCH), 5.36 (dd, 1H, H-3, J_{3,4} = 3.4 Hz), 4.97 (d, 1H, H-1), 4.60 (bd, 1H, H-4), 4.46 (dd, 1H, H-6a, J_{5,6a} = 1.3 Hz, J_{6a,6b} = 12.3 Hz), 4.10 (dd, 1H, H-6b, J_{5,6b} = 1.4 Hz), 3.77 (m, 1H, H-5); EIMS (m/z): 459 (M-PhS), 337, 105 (base).

Anal. Calcd for C₃₃H₂₈O₇S (568.64): C, 69.70; H, 4.96. Found: C, 69.57; H, 4.86.

Phenyl 2,3-Di-*O*-acetyl-1-thio-β-D-galactopyranoside (7a). A solution of 6a (703 mg, 1.60 mmol) in 80% HOAc (65 mL) was stirred for 2 h at 80 °C, then concentrated *in vacuo*. The residue was purified by flash chromatography (CH₂Cl₂: MeOH 30 : 1) to afford 7a (536 mg, 95%) as a colorless syrup: [α]_D +13.3° (c 0.23 CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 7.6-7.3 (m, 5H, C₆H₅), 5.32 (t, 1H, H-2, J_{1,2} = J_{2,3} = 9.9 Hz), 5.00 (dd, 1H, H-3, J_{3,4} = 2.9 Hz), 4.75 (d, 1H, H-1), 4.19 (dd, 1H, H-4, J_{4,5} = 0.7 Hz), 3.97 (dd, 1H, H-6a, J_{6a,6b} = 12.0 Hz, J_{5,6a} = 5.9 Hz), 3.88 (dd, 1H, H-6b, J_{5,6b} = 4.3 Hz), 3.66 (m, 1H, H-5); EIMS (m/z): 247 (M-PhS), 229, 187, 169, 145, 43 (base).

Phenyl 2,3-Di-*O*-benzoyl-1-thio-β-D-galactopyranoside (7b). A procedure similar to that for the preparation of 7a was employed. Compound 6b (1.0 g, 1.76 mmol) afforded 7b (673 mg, 80%): mp 183-184 °C; $[\alpha]_D$ +94.5° (*c* 1.6 CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.1-7.3 (m, 15H, 3C₆H₅), 5.80 (dd, 1H, H-2, J_{1,2} = 10.0 Hz, J_{2,3} = 9.9 Hz), 5.34 (dd, 1H, H-3, J_{3,4} = 2.9 Hz), 4.98 (d, 1H, H-1), 4.43 (bd, 1H, H-4), 4.05 (dd, 1H, H-6a, J_{5,6a} = 5.8 Hz, J_{6a,6b} = 11.9 Hz), 3.94 (dd, 1H, H-6b, J_{5,6b} = 3.8 Hz), 3.83 (m, 1H, H-5); EIMS (m/z): 371 (M-PhS), 365, 249, 105 (base).

Anal. Calcd for C₂₆H₂₄O₇S (480.53): C, 64.99; H, 5.03. Found: C, 64.52; H, 4.84.

Phenyl 2,3-Di-O-acetyl-4,6-O-[(R)-1-methoxycarbonylethylidene]-1-thio- β -D-galactopyranoside (8a). To a solution of 7a (400 mg, 1.12 mmol) in dry MeCN (3 mL) was added methyl pyruvate (250 mg, 2.45 mmol) and BF₃-OEt₂ (0.28 mL, 2.19 mmol). After being stirred for 5 h at rt under N₂, the mixture was quenched with saturated NaHCO₃ solution, then extracted with EtOAc. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. Flash chromatography of the residue (petroleum ether:EtOAc 8:1 \rightarrow 6:1 \rightarrow 4:1 \rightarrow 2:1) afforded 8a (146 mg, 30%) and recovered 7a (122 mg, 30%). 8a: mp 72-73 °C; $[\alpha]_D + 2.1^\circ$ (c 0.29 CH₂Cl₂); ¹H NMR

(300 MHz, CDCl₃) δ 7.6-7.3 (m, 5H, C₆H₅), 5.30 (t, 1H, H-2, J_{1,2} = J_{2,3} = 9.9 Hz), 4.85 (dd, 1H, H-3, J_{3,4} = 3.4 Hz), 4.64 (d, 1H, H-1), 4.32 (d, 1H, H-4), 4.12 (dd, 1H, H-6a, J_{6a,6b} = 12.8 Hz, J_{5,6a} = 2.5 Hz), 3.93 (dd, 1H, H-6b, J_{5,6b} = 1.7 Hz), 3.75 (s, 3H, CH₃O), 2.09, 2.07 (2s, 2×3H, 2CH₃CO), 1.51 (s, 3H, CH₃); EIMS (m/z): 381 (M-Ac), 331 (M-PhS), 271, 229, 201, 43 (base).

Phenyl 2,3-Di-*O*-benzoyl-4,6-*O*-[(*R*)-1-methoxycarbonylethylidene]-1-thio-β-D-galactopyranoside (8b). A procedure similar to that for the preparation of 8a was employed. Compound 7b (673 mg, 1.4 mmol) was treated with methyl pyruvate (286 mg, 2.8 mmol) and BF₃-OEt₂ (0.35 mL, 2.8 mmol) to afford 8b (505 mg, 64%): mp 81-83 °C; [α]_D +45.4° (c 0.6 CHCl₃) [Lit¹² +44° (c 1.0 CHCl₃)]; ¹H NMR (300 MHz, CDCl₃) δ 8.1-7.3 (m, 15H, 3C₆H₅), 5.77 (dd, 1H, H-2, J_{1,2} = 9.8 Hz, J_{2,3} = 10.0 Hz), 5.22 (dd, 1H, H-3, J_{3,4} = 3.5 Hz), 4.92 (d, 1H, H-1), 4.57 (dd, 1H, H-4), 4.22 (dd, 1H, H-6a, J_{6a,6b} = 12.4 Hz, J_{5,6a} = 1.5 Hz), 4.04 (dd, 1H, H-6b, J_{5,6b} = 1.7 Hz), 3.66 (s, 3H, CH₃O), 3.45 (bs, H-5), 1.54 (s, 3H, CH₃); EIMS (m/z): 505, 455, 333, 105 (base).

Anal. Calcd for C₃₀H₂₈O₉S (564.60): C, 63.82; H, 5.00. Found: C, 63.68; H, 4.99.

Phenyl 4,6-*O*-Benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (11). To a solution of 9¹⁸(9.0 g, 18.9 mmol) in dry CH₂Cl₂ (75 mL) was added PhSH (2.52 mL, 24.5 mmol) and BF₃-OEt₂ (23.2 mL, 184 mmol). After being stirred for 15 h at rt under N₂, the reaction was quenched with saturated NaHCO₃ solution (200 mL), then extracted with CH₂Cl₂. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. Flash chromatography of the residue (petroleum ether:EtOAc 4:1 \rightarrow 3:1 \rightarrow 2:1) afforded 10 (7.5 g, 75%).

To a solution of 10 (2.76 g, 5.24 mmol) in MeOH (100 mL) and CH₂Cl₂ (100 mL) was added a catalytic amount of NaOMe. The solution was stirred for 2 h at rt, then neutralized with Dowex-50WX8 (H⁺ form), filtered, and concentrated *in vacuo*. The residue was dissolved in dry MeCN (20 mL), then PhCH(OMe)₂ (3.1 mL) was added followed by TsOH-H₂O (100 mg). After being stirred for 15 h at rt, the mixture was quenched with Et₃N, and extracted with EtOAc. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. Flash chromatography of the residue (petroleum ether:EtOAc 3:1) afforded 11 (1.89 g, 80%) as a white syrup: [α]_D +41.2° (c 0.8 CH₂Cl₂) [Lit²³ +34.2° (c 1.3 CHCl₃); ¹H NMR (300

MHz, CDCl₃) δ 8.0-7.2 (m, 14 H, 2C₆H₅, C₆H₄), 5.70 (d, 1H, H-1, J_{1,2} = 10.4 Hz), 5.57 (s, 1H, PhCH), 4.64 (dd, 1H, H-3, J_{2,3} = 9.0 Hz, J_{3,4} = 10.0 Hz), 4.41 (dd, 1H, H-6a, J_{6a,6b} = 10.1 Hz, J_{5,6a} = 4.5 Hz), 4.34 (dd, 1H, H-6b, J_{5,6b} = 9.4 Hz), 3.83 (dd, 1H, H-4, J_{4,5} = 9.9 Hz), 3.72 (ddd, 1H, H-5), 3.61 (dd, 1H, H-2); EIMS (m/z): 380 (M-PhS), 362, 316, 256, 149, 91 (base).

Phenyl 3-*O*-Benzyl-4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (12). To a solution of 11 (900 mg, 1.94 mmol) in dry DMF (9 mL) was added NaH (110 mg, 60% in mineral oil, 2.75 mmol). After being stirred for 30 min at 0 °C under N₂, BnBr (0.26 mL, 2.17 mmol) was added dropwise, and the reaction mixture was then stirred overnight at rt. The mixture was diluted with MeOH (2 mL), then extracted with EtOAc. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. Flash chromatography of the residue (petroleum ether:EtOAc 5:1→3:1) afforded 12²³ as a colorless syrup (1.0 g, 84%) and recovered 11 (100 mg). 12: [α]_D +73.2° (c 4.2 CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 8.0-6.8 (m, 19H, 3C₆H₅, C₆H₄), 5.62 (s, 1H, PhCH), 5.61 (d, 1H, H-1, J_{1,2} = 10.4 Hz), 4.78 and 4.49 (AB, 2H, PhCH₂, J = 12.3 Hz), 4.5-4.4 (m, 2H, H-3, H-6a), 4.29 (t, 1H, H-6b, J_{6a,6b} = 10.1 Hz, J_{5,6b} = 10.1 Hz), 3.86 (dd, 1H, H-4, J_{3,4} = 9.9 Hz, J_{4,5} = 9.5 Hz), 3.80 (dd,1H, H-2, J_{2,3} = 8.8 Hz), 3.71 (ddd, 1H, H-5, J_{5,6a} = 4.6 Hz); EIMS (m/z): 470 (M-PhS), 362, 256, 91 (base).

Phenyl 3,6-Di-O-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (13). A mixture of 12 (888 mg, 1.53 mmol) and 4Å MS (1 g) in dry THF (10 mL) was stirred for 3 h, then NaBH₃CN (960 mg) was added followed by addition of saturated HCl in dry Et₂O until gas evolution ceased. After 5 min the suspension was filtered and diluted with EtOAc. The organic layer was washed with brine, dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography to afford 13^{16,24} (888 mg, 100%) as a colorless syrup: [α]_D +63.3° (c 5.2 CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 7.9-6.8 (m, 19H, 3C₆H₅, C₆H₄), 5.55 (d, 1H, H-1, J_{1,2} = 10.2 Hz), 4.73 and 4.53 (AB, 2H, PhCH₂, J = 12.1Hz), 4.62 and 4.57 (AB, 2H, PhCH₂, J = 11.8 Hz), 4.3-4.2 (m, 2H, H-6a, H-6b), 3.9-3.8 (m, 3H, H-2, H-3, H-4), 3.70 (m, 1H, H-5); EIMS (m/z): 472 (M-PhS), 364, 226, 91 (base).

Phenyl 4-*O*-Acetyl-3,6-di-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (14). A solution of 13 (1.041 g, 1.79 mmol) in dry pyridine (2 mL) and Ac₂O (2 mL) was stirred overnight. The solution was quenched with MeOH, then diluted with EtOAc. The organic layer was washed with 5% HCl solution, saturated NaHCO₃ solution, and brine respectively, and then dried over anhyd MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (petroleum ether:EtOAc 4:1) to afford $14^{16,24}$ (1.04 g, 100%) as a white syrup: [α]_D +96.4° (*c* 3.9 CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 7.9-6.8 (m, 19H, 3C₆H₅, C₆H₄), 5.55 (d, 1H, H-1, J_{1,2} = 10.4 Hz), 5.12 (dd, 1H, H-4, J_{3,4} = 9.2 Hz, J_{4,5} = 9.7 Hz), 4.60 and 4.31 (AB, 2H, PhCH₂, J = 12.0 Hz), 4.54 (s, 2H, PhCH₂), 4.45 (dd, 1H, H-3, J_{2,3} = 10.0 Hz), 4.32 (dd, 1H, H-2), 3.81 (m, 1H, H-5), 3.68-3.60 (m, 2H, H-6a, H-6b), 1.96 (s, 3H, CH₃CO); EIMS (*m/z*): 530, 454, 91.

Anal. Calcd for $C_{36}H_{33}NO_7S$ (623.72): C, 69.33; H, 5.33. Found: C, 69.37; H, 5.28.

2,4-Di-O-benzyl-3-O-(4-O-acetyl-3,6-di-O-benzyl-2-deoxy-2-phthal-Methyl imido- β -D-glucopyranosyl)- β -L-fucopyranoside (16). A suspension of donor 14 (1.09 g, 1.73 mmol), acceptor 15 (517 mg, 1.44 mmol) and 4ÅMS (0.5 g) in dry CH₂Cl₂ (10 mL) was stirred for 1 h at rt under Ar, then cooled to -30 °C, NIS (520 mg, 2.31 mmol) was added followed by immediate addition of a solution of AgOTf (130 mg, 0.51 mmol) in dry PhMe (8 mL). The mixture was stirred for 15 min, diluted with saturated Na₂S₂O₃ solution (2mL), filtered, and then diluted with EtOAc. The organic layer was washed with saturated Na₂S₂O₃ solution and brine respectively, then dried over anhyd MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography (petroleum ether:EtOAc 6:1 \rightarrow 5:1 \rightarrow 3:1) to afford 16 (912 mg, 72%): mp 55-56 °C; [α]_D -15.5° (c 0.4 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃, DQFCOSY, TOCSY) δ 7.5-6.8 (m, 24H, $4C_6H_5$, C_6H_4), 5.48 (d, 1H, H-1', $J_{1',2'} = 8.2$ Hz), 5.16 (dd, 1H, H-4', $J_{3',4'} = 9.6$ Hz, $J_{4'.5'} = 9.2 \text{ Hz}$), 4.80 and 4.72 (AB, 2H, PhCH₂, J = 11.3 Hz), 4.60 and 4.31 (AB, 2H, PhCH₂, J= 12.1 Hz), 4.51 and 4.40 (AB, 2H, PhCH₂, J= 11.8 Hz), 4.43 (dd, 1H, H-3', $J_{2',3'} = 9.0 \text{ Hz}$), 4.37 (dd, 1H, H-2'), 4.36 and 4.10 (AB, 2H, PhCH₂, J= 11.2 Hz), 4.12 (d, 1H, H-1, $J_{1,2} = 7.5$ Hz), 3.90 (dd, 1H, H-3, $J_{2,3} = 9.7$ Hz, $J_{3,4} = 2.5$ Hz), 3.76 (ddd, 1H, H-5', $J_{5',6a'} = 2.0 \text{ Hz}$, $J_{5',6b'} = 6.0 \text{ Hz}$), 3.595 (dd, 1H, H-2), 3.586 (dd, 1H, H-6a', $J_{6a',6b'} =$ 10.8 Hz), 3.51 (dd, 1H, H-6b'), 3.46 (s, 3H, CH₃O), 3.4-3.3 (m, 2H, H-4, H-5), 1.95 (s, 3H, CH₃CO), 1.115 (d, 3H, CH₃-6, J_{5,6} = 6.4 Hz). FABMS (*m/z*): 872 (M+1), 871 (M).

Anal. Calcd for $C_{51}H_{53}NO_{12}$ (871.99): C, 70.25; H, 6.13; N, 1.61. Found: C, 70.44; H, 6.25; N, 1.68.

Methyl 2,4-Di-O-benzyl-3-O-(2-acetamido-3,6-di-O-benzyl-2-deoxy-β-D-glucopyranosyl)-β-L-fucopyranoside (17). A solution of 16 (600 mg, 0.688 mmol) and hydrazine monohydrate (1 mL) in 95% EtOH (20 mL) was refluxed for 10 h, then concentrated in vacuo. Traces of water and ethanol were removed by coevaporating with toluene several times. The residue was stirred overnight in dry pyridine (4 mL) and Ac₂O (4 mL), then the solvent was removed by coevaporating with toluene several times. The resulting crude product was dissolved in MeOH (10 mL) and CH₂Cl₂ (10 mL), then a catalytic amount of NaOMe was added. The solution was stirred overnight, neutralized with Dowex-50WX8 (H⁺ form), filtered, and concentrated in vacuo. The residue was purified by flash chromatography (petroleum ether:EtOAc 1.5:1) to give 17 (377 mg, 74%): mp 162-164 °C; $[\alpha]_D$ -106.6° (c 0.63 CH₂Cl₂); ¹H NMR (300 MHz, CDCl₃) δ 7.5-7.1 (m, 20H, $4C_6H_5$), 5.43 (d, 1H, NHAc, J = 6.9 Hz), 5.28 (d, 1H, H-1', $J_{1'.2'} = 8.0$ Hz), 4.93 and 4.79 (AB, 2H, PhCH₂, J= 11.6 Hz), 4.85 and 4.62 (AB, 2H, PhCH₂, J= 11.8 Hz), 4.70 and 4.63 (AB, 2H, PhCH₂, J= 11.7 Hz), 4.48 (s, 2H, PhCH₂), 4.22 (dd, 1H, H-4', $J_{3,4} = 9.0 \text{ Hz}, J_{4,5} = 9.9 \text{ Hz}, 4.20 \text{ (d, 1H, H-1, } J_{1,2} = 7.5 \text{ Hz}), 3.85 \text{ (dd, 1H, H-3, } J_{2,3} = 7.5 \text{ Hz}$ 9.9 Hz, $J_{3,4} = 2.9$ Hz), 3.68 (dd, 1H, H-6a', $J_{5',6a'} = 2.5$ Hz, $J_{6a',6b'} = 10.0$ Hz), 3.65-3.64 (m, 2H, H-6b', H-2), 3.60-3.51 (m, 3H, H-2', H-3', H-4'), 3.50 (s, 3H, CH₃O), 3.46 (q, 1H, H-5, $J_{5,6} = 6.4$ Hz), 3.12 (ddd, 1H, H-5', $J_{5',6b'} = 7.5$ Hz), 1.49 (s, 3H, CH₃CO), 1.17 (d, 3H, CH_3 -6); FABMS (m/z): 765 (M+1+Na), 760 (M+1+H₂O), 742 (M+1), 384.

Anal. Calcd for $C_{43}H_{51}NO_{10}$ (741.87): C, 69.62; H, 6.93; N, 1.89. Found: C, 69.36; H, 6.84; N, 1.60.

Methyl 2,4-Di-O-benzyl-3-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-β-L-fucopyranoside (18). To a solution of 16 (190 mg, 0.218 mmol) in MeOH (2 mL) and CH₂Cl₂ (2 mL) was added a catalytic amount of NaOMe. The solution was stirred for 1 h at rt, neutralized with Dowex-50WX8 (H⁺ form), filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (petroleum ether:EtOAc 3:1) to afford 18 (180 mg, 100%): mp 62-63 °C; [α]_D -34.64° (c 0.6 CHCl₃);

¹H NMR (300 MHz, CDCl₃) δ 7.6-6.7 (m, 24H, 4C₆H₅, C₆H₄), 5.46 (d, 1H, H-1', $J_{1',2'}$ = 7.8 Hz), 4.77 and 4.53 (AB, 2H, PhCH₂, J = 14.5 Hz), 4.75 (m, 2H, PhCH₂), 4.50 (s, 2H, PhCH₂), 4.39 and 4.05 (AB, 2H, PhCH₂, J = 11.2 Hz), 4.24 (m, 2H, H-2', H-3'), 4.16 (d, 1H, H-1, $J_{1,2}$ = 7.4 H), 3.86 (dd, 1H, H-4', $J_{3',4'}$ = 7.3 Hz, $J_{4',5'}$ = 8.7 Hz), 3.85 (dd, 1H, H-3, $J_{2,3}$ = 9.7 Hz, $J_{3,4}$ = 2.4 Hz), 3.70-3.56 (m, 4H, H-2, H-5', H-6a', H-6b'), 3.45 (s, 3H, CH₃O), 3.39 (q, 1H, H-5, $J_{5,6}$ = 6.4 Hz), 3.33 (bd, 1H, H-4), 1.10 (d, 3H, CH₃-6); FABMS (m/z): 852 (M+Na), 830 (M+1), 829 (M).

Anal. Calcd for $C_{49}H_{51}NO_{11}$ (829.94): C, 70.91; H, 6.19; N, 1.69. Found: C, 70.51; H, 6.10; N, 1.91.

Methyl 2,3-Di-O-benzoyl-4,6-O-[(R)-1-methoxycarbonylethylidene]-β-Dgalactopyranosyl- $(1\rightarrow 4)$ -3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1→3)-2,4-di-O-benzyl-β-L-fucopyranoside (19). A suspension of 8b (150 mg, 0.266 mmol), 18 (110 mg, 0.133 mmol), and 4Å MS (0.1 g) in dry CH₂Cl₂ (3 mL) was stirred for 1 h at rt under Ar, then cooled to -20 °C. NIS (45 mg, 0.2 mmol) was added followed by immediate addition of a solution of AgOTf (17 mg, 0.066 mmol) in dry PhMe (1 mL). The reaction mixture was stirred for 30 min, quenched with saturated Na₂S₂O₃ solution (2 mL), filtered, and then diluted with EtOAc. The organic layer was washed with saturated Na₂S₂O₃ solution and brine respectively, dried over anhyd MgSO₄, filtered, and concentrated in vacuo. The residue was purified by flash chromatography (petroleum ether:EtOAc 6:1 \rightarrow 5:1 \rightarrow 3:1) to afford 19 (120 mg, 71%): mp 94-95 °C; ; [α]_D -35.7° (c 0.5 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃, DOFCOSY, TOCSY) δ 8.0-6.7 (m. 34H, $6C_6H_5$, C_6H_4), 5.80 (dd, 1H, H-2", $J_{1"2"} = 8.1$ Hz, $J_{2"3"} = 10.2$ Hz), 5.33 (d, 1H, H-1', $J_{1',2'} = 8.0$ Hz), 5.05 (dd, 1H, H-3'', $J_{3'',4''} = 3.6$ Hz), 5.02 and 4.55 (AB, 2H, $PhCH_2$, J = 12.3 Hz), 4.86 (d, 1H, H-1"), 4.78 and 4.69 (AB, 2H, $PhCH_2$, J = 11.8 Hz), 4.59 and 4.31 (AB, 2H, PhCH₂, J = 12.2 Hz), 4.44 (d, 1H, H-4"), 4.39 and 4.03 (AB, 2H, PhCH₂, J = 11.3 Hz), 4.26 (dd, 1H, H-2'), 4.20 (m, 1H, H-5'), 4.16 (m, 1H, H-6a''), 4.12 (d, 1H, H-1, $J_{1,2} = 7.5$ Hz), 3.95 (d, 1H, H-6b'', $J_{6a'',6b''} = 11.3$ Hz), 3.86 (dd, 1H, H-4', $J_{3',4'} = J_{4',5'} = 9.1 \text{ Hz}$), 3.77 (dd, 1H, H-3, $J_{2,3} = 9.7 \text{ Hz}$, $J_{3,4} = 2.6 \text{ Hz}$), 3.66 (dd, 1H, H-3', $J_{2',3'} = 7.0 \text{ Hz}$), 3.63 (s, 3H, CH₃O), 3.57 (dd, 1H, H-2), 3.42 (s, 3H, CH₃O), 3.35-3.27 (m, 4H, H-5, H-5", H-6a', H-6b"), 3.24 (d, 1H, H-4), 1.43 (s, 3H, CH₃), 1.05 (d, 3H,

CH₃-6, $J_{5,6} = 6.4$ Hz); FABMS (m/z): 1284 (M+1), 456, 105 (base); IR (KBr, cm⁻¹): 2935, 2871, 1777, 1715, 1602, 1585, 1110, 1092, 1071.

Anal. Calcd for $C_{73}H_{73}NO_{20}$ (1284.37): C, 68.27; H, 5.73; N, 1.09. Found: C, 68.34; H, 5.96; N, 1.19.

Methyl 4,6-O-[(R)-1-Carboxyethylidene]- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl- $(1\rightarrow 3)$ - β -L-fucopyranoside (1b). To a solution of 19 (62 mg, 0.048 mmol) in iso-PrOH (5 mL) and H₂O (0.8 mL) was added NaBH₄ (18 mg, 0.26 mmol). After being stirred overnight at rt, the solution was acidified by AcOH to pH 5, then heated to 50 °C for 5 h. The solution was concentrated in vacuo and traces of water and HOAc were removed by coevaporating with toluene several times. The residue was dissolved in dry pyridine (1 mL) and Ac₂O (1 mL). After being stirred overnight, the mixture was diluted with MeOH (2 mL) then with EtOAc. The organic layer was washed with 5% HCl solution, saturated NaHCO3, and brine respectively, and then dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The residue was debenzoylated with a catalytic amount of NaOMe in MeOH (2 mL) at rt for 2 h, saponified with aq NaOH (13 mg, 1mL H₂O and 1mL MeOH, 2 h at rt). The mixture was neutralized with Dowex-50WX8 (H⁺ form), filtered, and concentrated in vacuo. The residue was hydrogenolyzed with a catalytic amount of 10% Pd/C (20 mg, 1 atm H₂ for 2 days), then the suspension was filtered and concentrated in vacuo. The crude product was purified by flash chromatography (CH₂Cl₂:MeOH $3:1\rightarrow 2:1\rightarrow 1:1$) to give 1b (14 mg, 44% from 19): $[\alpha]_D$ -58° (c 0.3 H₂O); ¹H NMR (600 MHz, D₂O, DQFCOSY) δ 4.86 (d, 1H, H-1', $J_{1',2'} = 8.5$ Hz), 4.53 (d, 1H, H-1'', $J_{1'',2''} = 7.8$ Hz), 4.36 (d, 1H, H-1, $J_{1,2} = 7.8$ Hz), 3.58 (s, 3H, CH₃O), 1.93 (s, 3H, CH₃CO), 1.42 (s, 3H, CH₃), 1.27 (d, 3H, CH₃-6); ESIMS (m/z): 699 (M+K-1), 662 (M+1).

ACKNOWLEDGMENT

This work was supported by the State Science and Technology Committee of China. We thank Prof. Houming Wu for kind assistance in recording NMR spectra.

REFERENCES

1. P. S. Galtsoff, Biol. Bull., 57, 250 (1929).

- 2. H. V. Wilson, J. Exp. Zool., 5, 245 (1907).
- 3. D. Spillmann, K. Hard, J. Thomas-Oates, J. F. G. Vliegenthart, G. Misevic, M. M. Burger and J. Finne, J. Biol. Chem., 268, 13378 (1993).
- 4. D. Spillmann, J. Thomas-Oates, J. A. von Kuik, J. F. G. Vliegenthart, G. Misevic, M. M. Burger and J. Finne, J. Biol. Chem., 270, 5089 (1995).
- 5. T. Humphreys, Dev. Biol., 8, 27 (1963).
- 6. J. E. Jumblatt, V. Schlup and M. M. Burger, Biochemistry, 19, 1038 (1980).
- 7. G. N. Misevic, J. Finne and M. M. Burger, J. Biol. Chem., 262, 5870 (1987).
- 8. G. N. Misevic and M. M. Burger, J. Biol. Chem., 265, 20577 (1990).
- 9. Z. W. Guo, S. J. Deng and Y. Z. Hui, J. Carbohydr. Chem., 15, 965 (1996).
- 10. T. Ziegler, Liebigs Ann. Chem., 949 (1995).
- 11. N. Khiar and M. M. Lomas, J. Org. Chem., 60, 7017 (1995).
- 12. T. Ziegler, E. Eckhardt and G. Herold, Tetrahedron Lett., 33, 4413 (1992).
- 13. T. Ziegler, E. Eckhardt, J. Strayle and H. Herzog, Carbohydr. Res., 253, 167 (1994).
- 14. P. J. Garegg and H. Hultberg, Carbohydr. Res., 93, c10 (1981).
- 15. K. Takeo, K. Nagayoshi, K. Nishimura and S. Kitamura, *J. Carbohydr. Chem.*, 13, 1159 (1994).
- 16. O. Kanie, Y. Ito and T. Ogawa, J. Am. Chem. Soc., 116, 12073 (1994).
- 17. J. O. Osby, M. G. Martin and B. Ganem, *Tetrahedron Lett.*, 25, 2093 (1984).
- 18. F. Dasgupta and P. J. Garegg, J. Carbohydr. Chem., 7, 701 (1988).
- 19. A. B. Landge, T. R. Ingle and S. L. Bose, Indian J. Chem., 7, 1200 (1969).
- 20. A. Liptak, I. Jordal, J. Harangi and P. Nanasi, Acta Chim. Hung., 113, 415 (1983).
- 21. P. de Pouilly, A. Chenede, J-M Mallet and P Sinay, Bull. Soc. Chim. Fr., 256 (1993).
- 22. T. Ziegler, E. Eckhardt and G. Herold, Liebigs Ann. Chem., 441 (1992).
- 23. R. K. Jain and K. L. Matta, Carbohydr. Res., 226, 91 (1992).
- 24. H. Paulsen, Angew. Chem., Int. Ed. Engl., 34, 1432 (1995).